

Absolute Stereochemistry of Isosaraine-1 and Isosaraine-2

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Abstract: The absolute stereochemistry of the quinolizidone systems present in the marine macrocyclic alkaloids isosaraine-1 (3) and isosaraine-2 (4), minor metabolites of the Mediterranean sponge Reniera sarai, has been established by applying advanced Mosher's method to the alcohol derivatives (5 and 6).

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The sponge *Reniera sarai*, order Haplosclerida, is widely present in the Bay of Naples, Italy. Its metabolic pattern is heavily characterized by a series of complex polycyclic diamine alkaloids, saraines, which possess interesting phase transfer catalytic properties² and biological activities.³

Up to now, nine macrocyclic alkaloids, saraines A-C, 4-6 saraines 1-3, 2,7 and isosaraines 1-3, 7-9 have been isolated and structurally characterized by our group. Two different skeletons were exhibited by saraines A-C (examplified by saraine-A, 1) and saraines 1-3 (examplified by saraine-2, 2) whereas isosaraines -1 (3), -2 (4), and -37 were characterized by having the same framework of saraines 1-3 but displaying inverted stereochemistry at the chiral centers C-1, C-2, and C-9. Very recently, the absolute stereochemistry of the quinolizidone moiety of saraines -1 and -2 have been established, while the absolute configurations of isosaraines remain undefined. In particular, the stereochemical relationships between saraines 1-3 and isosaraines 1-3 are extremely relevant in the light of the recent paper 10 reporting the total synthesis of racemic petrosin (7) identical to the natural compound. The authors stated that being uncommon for natural products to be biosynthesized in racemic form, the presence in Petrosia seriata of racemic alkaloids could be due to some post-biosynthetic equilibrations. Of course, the chiral centers α to the carbonyl can be very easily epimerized whereas the more remote stereocenters might epimerize by retro-Mannich-Mannich and immonium ion-enamine equilibria (Scheme 1). According to this hypothesis, isosaraines 1-3 could biosynthetically derive from saraines 1-3. Because of this, it is very relevant to establish their absolute stereostructure. For a better understanding of both chemistry and biogenetic origin of saraines, we have recently studied the absolute stereochemistry of isosaraines. This paper describes the elucidation of the absolute stereochemistry of isosaraines -1 (3) and -2 (4).

Isosaraine-1 (3) was first subjected to reduction with NaBH₄ to afford the derivative 5 as a single product. The structure of 5^{11} was fully analyzed by detailed NMR analysis confirming the relative configurations at C-1, C-2, C-9 and C-10. Moreover, 1 H- and 13 C-NMR spectra allowed to assign an oxymethine ($\delta_{\rm H}$ 3.60, $\delta_{\rm C}$ 72.5) at C-8 suggesting that 5 was the 8-hydroxy-isosaraine-1. The equatorial orientation of the hydroxy group was determined on the basis of decoupling experiments. In fact, irradiation at

H-9 ($\delta_{\rm H}$ 1.88, $\delta_{\rm C}$ 42.6) simplified the H-8 multiplet to a double doublet exhibiting axial-axial (J = 9.9 Hz) and axial-equatorial (J = 4.3 Hz) couplings with the vicinal H₂-7 protons.

Treatment of **5** with R-(-)- and S-(+)- α -methoxy- α -(trifluromethyl)phenylacetyl chloride (MTPA-Cl) yielded the corresponding S (**5s**)- and R (**5r**)-MTPA esters, respectively.

Table 1. Selected ¹H-NMR Chemical Shifts^a for the MTPA Esters of the Reductive Products of Isosaraines 1-2 (**5** and **6**) and $\Delta\delta$ ($\delta_{S-MTPA \ ester}$ - $\delta_{R-MTPA \ ester}$)^b

Н	5 S	5 R	$\Delta \delta$	6 S	6 R	$\Delta \delta$
4a	2.67	2.66	+5	2.69	2.69	0
4b	1.85	1.85	0	1.87	1.87	0
6a	2.81	2.78	+15	2.81	2.79	+10
6b	2.00	1.98	+10	2.00	2.00	0
7a	1.86	1.76	+50	1.85	1.76	+45
7b	1.80	1.76	+20	1.81	1.76	+25
8	4.93	4.93	0	4.93	4.92	+5
9	1.94	1.99	-25	1.98	2.02	-20
10	1.81	1.83	-10	1.70	1.72	-10
1	1.62	1.63	-5	1.56	1.58	-10
2	1.50	1.51	-5	1.92	1.92	0
a_1	1.59	1.53	-30	1.52	1.56	-20
a_2	1.59	1.53	-30	1.27	1.32	-25
b_1^2	1.19	1.22	-15	1.12	1.20	-40
b_2	1.19	1.22	-15	0.89	1.00	-55

^a Bruker AMX 500 MHz; CDCl₃; δ values referred to CHCl₃ (δ 7.26 ppm); The assignments were aided by $^{1}\text{H-}^{1}\text{H}$ COSY, TOCSY and $^{1}\text{H-}^{13}\text{C}$ HETCOR.

b $\Delta \delta$ values are given in Hz.

The ¹H-NMR chemical shifts of **5s** and **5R** were carefully assigned by extensive interpretation of ¹H¹H COSY, TOCSY, and ¹H-¹³C HETCOR spectra. The proton chemical shift differences ($\Delta\delta = \delta_{S-MTPA-ester}$) for the protons near to the chiral center C-8 are listed in **Table 1**.

Analogously with the Mosher esters of equatorial 8-hydroxy-saraine-2,⁷ positive $\Delta\delta$ values were observed for H₂-7 and H₂-6 which were located on the right side of the MTPA plane, whereas negative values were observed for H-9, H₂-a, H₂-b, H-1, and H-2 which were located on the left side of the MTPA plane. According to MTPA determination rule,¹² the absolute stereochemistry of C-8 was assigned as R. Consequently, the absolute configurations at C-1, C-2, C-9, and C-10 of isosaraine-2 were concluded to be R, R, S, and R, respectively. Of course, it is impossible to determine the absolute stereochemistry at C-3', joining point of two independent heterocyclic systems.

In a similar manner, isosaraine-2 (4) was reduced to 6^{13} which was subsequently esterified by reacting with $R(\cdot)$ - and S(+)-MTPA chloride affording the pair of Mosher esters $6\mathbf{S}$ and $6\mathbf{R}$. The $\Delta\delta$ values $[\Delta\delta=\delta_S(6\mathbf{S})-\delta_R(6\mathbf{R})]$ of the protons near the chiral center C-8, as reported in **Table 1**, were very similar to those observed for Mosher esters ($5\mathbf{S}$ and $5\mathbf{R}$) of 8-hydroxy-isosaraine-1 (5) leading to the same assignments 1R, 2R, 8R, 9S, 10R for all the chiral centers of the quinolizidine moiety. The limited amounts of isosaraine-3 prevented from establishing its absolute stereochemistry by using the same procedure as mentioned above. However, considering that it is a superior homologue of 3 and 4, it probably displays the same absolute configuration (1R 2R, 9S, 10R).

Plausible biosyntetic pathway of saraines 1-3 and isosaraines 1-3 according to ref. 10

On the basis of the knowledge of both absolute stereochemistry of saraines 1-2 and isosaraines 1-2, it is fascinating to suggest a more insightful biogenetic pathway for them. The hypothesis proposed by Heathcock *et al* for petrosins 10 can, as shown in **Scheme 1**, justify the epimerization of the stereocenters C-1 and C-9 that might occur by retro-Mannich-Mannich and immonium ion-enamine equilibria while the configuration of the more remote stereocenter C-2 might be controlled, in a subsequent step when there is the coupling between C-2 and C-3', by the steric bulk of the alkyl chain at C-1 and the tetrahydropyridine ring at C-2. As a consequence, a *trans* orientation of the substituents at C-1 and C-2 is favoured in both saraines 1-3 and, with inverted configuration, isosaraines 1-3.

Acknowledgments: The NMR and the mass spectra were respectively obtained from the "ICMIB-NMR Service" and from the "Servizio di Spettrometria di Massa del CNR e dell'Università di Napoli". The staff of both services is gratefully acknowledged. The authors thank Mr. R. Turco for technical assistance. This research was supported by the C.N.R. strategic project "Tecnologie Chimiche Innovative". Dr Y.-W. Guo thanks the Ministry of Foreign Affairs of Italy for the financial support in the frame of the Italian-Chinese Scientific and Technological Cooperation. VIII program-Basic Science: "Chemical Mediators in Marine Invertebrates".

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- 11. Spectral data of equatorial 8-hydroxy-isosaraine-1 (5): Amorphous powder, $[\alpha]_D$ -27.9° (c 0.46, CHCl₃); EIMS, m/z (%): 468 (M⁺, 100), 329 (35); ¹H-NMR (CDCl₃) δ (ppm): 1.65 (m, H-1), 1.49 (m, H-2), 1.68, 1.54 (m, H₂-3), 2.70 (m, H-4a), 1.84 (m, H-4b), 2.78 (m, H-6a), 1.92 (m, H-6b), 1.67, 1.62 (m, H₂-7), 3.62 (ddd, J=9.9, 6.0, 5.1 Hz, H-8_{ax}), 1.88 (m, H-9eq), 1.73 (m, H-10ax), 2.72 (br d, J=10.5 Hz, H-2'_{eq}), 2.29 (m, H-2'_{ax}), 2.16 (m, H-3'), 5.46 (br s, H-4'), 3.14 (d, J=16.5 Hz, H-6'a), 2.40 (d, J=16.5 Hz, H-6'b), 1.32, 1.26 (m, H₂-a), 1.16 (m, H₂-b), 1.37 (m, H₂-d), 1.53, 1.02 (m, H₂-e), 1.86, 1.76 (m, H₂-f), 2.32 (m, H₂-g), 1.50 (m, H₂-h), 2.10, 1.90 (m, H₂-i), 5.56 (m, H-j), 5.27 (m, H-k), 2.20, 2.05 (m, H₂-l), 1.43 (m, H₂-n), 2.39, 1.27 (m, H₂-q); ¹³C-NMR (CDCl₃) δ (ppm): 34.4 (d, C-1), 41.7 (d, C-2), 32.3 (t, C-3), 54.3 (t, C-4), 55.4 (t, C-6), 30.0 (t, C-7), 73.2 (d, C-8), 43.9 (d, C-9), 68.1 (d, C-10), 55.4 (t, C-2'), 42.5 (d, C-3'), 119.5 (d, C-4'), 137.8 (s, C-5'), 55.0 (t, C-6'), 24.8 (t, C-a), 29.7 (t, C-b), 25.8 (t, C-d), 24.2 (t, C-e), 33.8 (t, C-f), 58.0 (t, C-g), 28.5 (t, C-h), 26.0 (t, C-i), 129.4 (t, C-j), 130.8 (t, C-k), 24.9 (t, C-l), 27.7 (t, C-m), 30.9 (t, C-q).
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- 13. Spectral data of equatorial 8-hydroxy-isosaraine-2 (6): Amorphous powder, $[\alpha]_D$ -37.9° (*c* 1.2, CHCl₃); EIMS, *m/z* (%): 456 (M⁺, 100), 329 (32); ¹H-NMR (CDCl₃) δ (ppm): 1.73 (m, H-1), 1.55 (m, H-2), 1.70, 1.55 (m, H₂-3), 2.70 (m, H-4a), 1.83 (m, H-4b), 2.77 (br d, H-6a), 1.92 (m, H-6b), 1.68, 1.63 (m, H₂-7), 3.60 (ddd, *J*=9.9, 6.1, 4.1 Hz, H-8_{ax}), 1.89 (m, H-9eq), 1.70 (m, H-10ax), 2.53 (m, H-2'a), 2.69 (m, H-2'b), 2.29 (br s, H-3'), 5.50 (s, H-4'), 3.18 (d, *J*=16.5 Hz, H-6'a), 2.49 (m, H-6'b), 1.50, 1.37 (m, H₂-a), 1.24 (m, H₂-b), 1.57, 1.08 (m, H₂-e), 1.88, 1.85 (m, H₂-f), 2.56, 2.47 (m, H₂-g), 1.62, 1.43 (m, H₂-h), 1.38 (m, H₂-i), 1.26, 1.17 (m, H₂-o), 2.21, 2.17 (m, H₂-p); ¹³C-NMR (CDCl₃) δ (ppm): 34.1 (d, C-1), 38.6 (d, C-2), 31.9 (t, C-3), 54.5 (t, C-4), 55.3 (t, C-6), 30.5 (t, C-7), 73.2 (d, C-8), 42.6 (d, C-9), 66.8 (d, C-10), 55.0 (t, C-2'), 42.6 (d, C-3'), 119.8 (d, C-4'), 137.9 (s, C-5'), 54.2 (t, C-6'), 24.0 (t, C-a), 29.5 (t, C-b), 26.7 (t, C-e), 33.6 (t, C-f), 57.1 (t, C-g), 23.4 (t, C-h), 26.9 (t, C-i), 22.3 (t, C-o), 28.9 (t, C-p).